Enzymatic activity of fungi isolated from crops

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Abstract

Aim: To detect and assess the activity of extracellular hydrolytic enzymes and to find differences in enzymograms between fungi isolated from wheat and rye samples and grown on Czapek-Dox Broth and Sabouraud Dextrose Broth enriched with cereal (wheat or rye). Isolated strains were also classified in the scale of biosafety levels (BSL). **Material and methods:** The study used 23 strains of fungi cultured from samples of wheat and rye (grain, grain dust obtained during threshing and soil) collected in the Lublin region (eastern Poland). API ZYM test (bioMérieux) was carried out according to the manufacturer's instructions. Classification of BSL (Biosafety levels) was based on the current literature.

Results: High enzymatic activity was found in strains cultured in media containing 1% of wheat grain (*Bipolaris holmi, Penicillium decumbens*) and with an addition of 1% of rye grain (*Cladosporium herbarum, Aspergillus versicolor, Alternaria alternata*). The total number of enzymes varied depending on the type of media, and in most cases it was higher in the culture where an addition of cereal grains was used.

Conclusions: Isolated strains of fungi reveal differences in the profiles of the enzyme assay. It can be assumed that the substrate enriched in grains stimulate the higher activity of mold enzymes.

Key words: enzymatic activity, mold fungi, zymogram, biohazards.

Introduction

API ZYM test (bioMérieux) is a semiquantitative assay allowing to assess the presence and activity of 19 hydrolytic enzymes. The release of hydrolytic enzymes into the environment by dermatophytes, yeasts, and molds is an important component in the pathogenesis of infection. Hydrolases (esterases, sulfatases, glycosidases and peptidases) facilitate the degradation of keratin, and thus the penetration of the mycelium into tissues [1–3].

The isolated and identified strains were also assigned to specific classes of biosafety. The Classification of Biosafety Level (BSL) determines the safety scale of potentially pathogenic fungi for humans and animals by highlighting four hazard classes. The class BSL-1 represents saprophytes or plant pathogens causing a coincidental, superficial, non-invasive or benign threat [4]. However, this may cause non-infectious respiratory diseases of an allergic or immunotoxic nature [5]. Class BSL-2 species are characterized by a relatively high ability to survive in the tissues of vertebrates, and in patients with severe immune disorders can cause a deep, opportunistic infection. Pathogens belonging to the class of BSL-3 are potentially capable of causing severe and deep fungal infections in apparently healthy subjects [4].

Aim

The aim of the study was to detect and assess the activity of extracellular hydrolytic enzymes and to find differences in enzymograms between fungi isolated from wheat and rye samples and grown on Czapek-Dox Broth

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| Isolate | Medium | | | | | | | | Er | ızym | e act | ivity* | ,* * | | | | | | | |
|------------------------|----------------------------------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|-------------|--------|--------|--------|--------|--------|--------|--------|
| | | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 |
| Alternaria | Czapek-Dox | 0 | 2 | 3 | 0 | 2 | 1 | 0 | 4 | 0 | 1 | 1 | 2 | 0 | 0 | 1 | 5 | 5 | 0 | 0 |
| alternata | Czapek-Dox + wheat | 4 | 2 | 3 | 0 | 5 | 5 | 1 | 2 | 2 | 4 | 5 | 0 | 0 | 2 | 3 | 5 | 5 | 0 | 0 |
| | Sabouraud | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 1 | 0 | 0 | 1 | 2 | 0 | 0 | 1 | 5 | 2 | 0 | 0 |
| | Sabouraud + wheat | 5 | 4 | 4 | 0 | 4 | 3 | 1 | 0 | 1 | 5 | 5 | 0 | 3 | 3 | 4 | 5 | 5 | 1 | 1 |
| Aspergillus | Czapek-Dox | 1 | 2 | 3 | 1 | 1 | 1 | 1 | 1 | 1 | 5 | 4 | 5 | 3 | 0 | 2 | 3 | 5 | 1 | 0 |
| fumigatus | Czapek-Dox + wheat Sabouraud | 0 3 | 1 2 | 0 1 | 0 0 | 3 5 | 4 1 | 0 2 | 0 1 | 0 2 | 3 5 | 4 4 | 5 1 | 3 3 | 0 0 | 4 0 | 5 3 | 3 4 | 0 4 | 0 0 |
| | Sabouraud + wheat | 1 | 1 | 2 | 0 | 3 | 2 | 2 | 2 | 2 | 5 | 2 | 3 | 3 | 0 | 3 | 3 | 4 | 3 | 0 |
| Aspergillus | Czapek-Dox | 3 | 2 | 3 | 1 | 0 | 0 | 0 | 0 | 0 | 5 | 5 | 5 | 2 | 0 | 1 | 5 | 4 | 1 | 0 |
| glaucus | Czapek-Dox + wheat | 2 | 2 | 2 | 0 | 0 | 1 | 0 | 0 | 0 | 2 | 2 | 4 | 1 | 0 | 0 | 2 | 3 | 2 | 0 |
| 9 | Sabouraud | 5 | 3 | 4 | 0 | 4 | 3 | 3 | 3 | 3 | 5 | 5 | 5 | 4 | 4 | 0 | 4 | 4 | 4 | 0 |
| | Sabouraud + wheat | 3 | 2 | 2 | 0 | 1 | 2 | 0 | 0 | 1 | 2 | 3 | 3 | 5 | 0 | 3 | 5 | 5 | 2 | 0 |
| Aspergillus | Czapek-Dox | 2 | 1 | 2 | 1 | 0 | 0 | 0 | 3 | 2 | 3 | 5 | 5 | 1 | 0 | 1 | 3 | 3 | 3 | 2 |
| sydowii | Czapek-Dox + wheat | 5 | 2 | 2 | 0 | 5 | 2 | 2 | 2 | 2 | 5 | 5 | 5 | 0 | 0 | 0 | 3 | 3 | 3 | 0 |
| | Sabouraud | 2 | 1 | 3 | 1 | 1 | 2 | 2 | 3 | 2 | 4 | 5 | 1 | 1 | 0 | 0 | 2 | 3 | 1 | 1 |
| | Sabouraud + wheat | 5 | 1 | 2 | 0 | 3 | 2 | 2 | 1 | 0 | 5 | 3 | 2 | 0 | 2 | 0 | 3 | 3 | 3 | 0 |
| Aspergillus | Czapek-Dox | 0 | 2 | 1 | 1 | 0 | 1 | 1 | 1 | 0 | 2 | 1 | 0 | 0 | 0 | 1 | 3 | 4 | 0 | 0 |
| tamarii | Czapek-Dox + wheat | 1 | 2 | 1 | 0 | 1 | 0 | 0 | 1 | 1 | 2 | 2 | 0 | 0 | 0 | 2 | 4 | 4 | 0 | 0 |
| | Sabouraud | 2 | 2 | 2 | 0 | 2 | 0 | 0 | 0 | 0 | 1 | 2 | 2 | 1 | 1 | 2 | 3 | 3 | 0 | 0 |
| | Sabouraud + wheat | 3 | 2 | 0 | 0 | 3 | 2 | 0 | 1 | 0 | 2 | 2 | 2 | 3 | 3 | 3 | 4 | 3 | 0 | 0 |
| Aspergillus | Czapek-Dox | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 3 | 3 | 2 | 0 | 1 | 0 | 2 | 2 | 0 | 0 |
| terreus | Czapek-Dox + wheat Sabouraud | 3 1 | 2 1 | 0 0 | 0 0 | 4 2 | 1 2 | 0 1 | 0 0 | 0 0 | 4 1 | 4 1 | 5 2 | 1 0 | 3 0 | 2 0 | 4 4 | 3 4 | 4 2 | 0 0 |
| | Sabouraud + wheat | 4 | 1 | 0 | 0 | 4 | 2 | 1 | 1 | 0 | 4 | 1 3 | 2 4 | 0 | 3 | 1 | 4 | 4 | 2 | 0 |
| Aspergillus | Czapek-Dox | 3 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 4 | 1 | 1 | 0 | 0 | 0 | 5 | 2 | 0 | 0 |
| versicolor | Czapek-Dox + wheat | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 1 | 2 | 1 | 0 | 0 | 4 | 0 | 0 | 0 |
| Versicolor | Sabouraud | 3 | 1 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 4 | 1 | 4 | 0 | 0 | 2 | 4 | 4 | 3 | 0 |
| | Sabouraud + wheat | 2 | 1 | 0 | 0 | Ő | 0 | 0 | 0 | 0 | 2 | 5 | 1 | 1 | 0 | 0 | 2 | 0 | 0 | 0 |
| Bipolaris holmi | Czapek-Dox | 1 | 0 | 2 | 0 | 1 | 0 | 0 | 0 | 0 | 2 | 2 | 4 | 3 | 0 | 3 | 5 | 5 | 1 | 1 |
| F | Czapek-Dox + wheat | 3 | 2 | 2 | 2 | 5 | 2 | 2 | 3 | 0 | 2 | 3 | 2 | 2 | 2 | 3 | 5 | 5 | 2 | 2 |
| | Sabouraud | 4 | 3 | 3 | 1 | 4 | 2 | 3 | 5 | 1 | 5 | 5 | 0 | 4 | 0 | 3 | 5 | 5 | 1 | 0 |
| | Sabouraud + wheat | 5 | 3 | 2 | 2 | 5 | 2 | 0 | 3 | 0 | 3 | 3 | 2 | 5 | 5 | 2 | 5 | 5 | 2 | 2 |
| Exserohilum | Czapek-Dox | 4 | 3 | 3 | 1 | 5 | 5 | 5 | 1 | 1 | 4 | 5 | 2 | 1 | 1 | 3 | 5 | 4 | 1 | 0 |
| sp. | Czapek-Dox + wheat | 1 | 1 | 0 | 1 | 2 | 1 | 0 | 4 | 1 | 0 | 2 | 4 | 4 | 0 | 2 | 5 | 4 | 1 | 0 |
| | Sabouraud | 3 | 2 | 1 | 0 | 4 | 2 | 2 | 3 | 2 | 5 | 5 | 5 | 3 | 3 | 5 | 5 | 3 | 3 | 2 |
| | Sabouraud + wheat | 3 | 2 | 2 | 0 | 2 | 0 | 0 | 2 | 2 | 3 | 3 | 3 | 2 | 2 | 5 | 3 | 5 | 2 | 0 |
| Fusarium | Czapek-Dox | 4 | 1 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 2 | 2 | 2 | 1 | 0 | 0 | 3 | 3 | 1 | 0 |
| proliferatum | Czapek-Dox + wheat | 3 | 1 | 2 | 1 | 5 | 0 | 1 | 0 | 1 | 4 | 3 | 5 | 4 | 0 | 1 | 5 | 5 | 3 | 0 |
| | Sabouraud Sabouraud + wheat | 4 | 1 1 | 2 3 | 0 0 | 4 5 | 1 1 | 1 0 | 0 0 | 0 1 | 3 3 | 3 4 | 0 3 | 0 2 | 0 0 | 0 0 | 3 3 | 2 3 | 0 0 | 0 0 |
| Fueravium | | 3 | | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 3 | 4 | 2 | 0 | 0 | 0 | 2 | 3 | 0 | |
| Fusarium tricinctum | Czapek-Dox Czapek-Dox + wheat | 4 | 1 2 | 2 | 0 | 4 | 0 | 0 | 2 | 0 | 5 | 2 | 2 | 2 | 0 | 0 | 2 5 | 5 4 | 1 | 0 |
| incinctum | Sabouraud | 4 | 2 | 3 | 1 | 5 | 2 | 0 | 0 | 0 | 3 | 2 | 1 | 0 | 0 | 0 | 1 | 4 | 0 | 0 |
| | Sabouraud + wheat | 5 | 3 | 4 | 0 | 3 | 0 | 0 | 0 | 0 | 5 | 5 | 5 | Ő | 0 | 1 | 5 | 5 | 2 | 0 |
| Penicillium | Czapek-Dox | 0 | 2 | 1 | 0 | 3 | 0 | 0 | 0 | 0 | 5 | 4 | 4 | 5 | 5 | 4 | 5 | 4 | 1 | 0 |
| decumbens | Czapek-Dox + wheat | 1 | 2 | 2 | 0 | 2 | 0 | 0 | 0 | 0 | 4 | 3 | 4 | 5 | 5 | 4 | 5 | 4 | 2 | 0 |
| | Sabouraud | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 4 | 2 | 0 | 5 | 1 | 0 | 5 | 5 | 0 | 0 |
| | Sabouraud + wheat | 3 | 2 | 2 | 2 | 3 | 2 | 0 | 0 | 0 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 4 | 4 | 0 |
| Penicillium | Czapek-Dox | 0 | 1 | 0 | 0 | 2 | 0 | 1 | 0 | 0 | 2 | 3 | 2 | 4 | 1 | 1 | 3 | 3 | 0 | 0 |
| expansum | Czapek-Dox + wheat | 0 | 1 | 2 | 0 | 2 | 0 | 1 | 0 | 0 | 3 | 3 | 3 | 5 | 4 | 3 | 4 | 4 | 0 | 0 |
| | Sabouraud | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 2 | 0 | 0 | 0 | 0 | 1 | 3 | 0 | 0 |
| | Sabouraud + wheat | 1 | 2 | 3 | 1 | 4 | 2 | 0 | 0 | 0 | 5 | 5 | 5 | 4 | 5 | 4 | 4 | 4 | 2 | 0 |
| Scopulariopsis | Czapek-Dox | 0 | 2 | 5 | 2 | 3 | 5 | 2 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 |
| brevicaulis | Czapek-Dox + wheat | 1 | 2 | 4 | 3 | 4 | 5 | 1 | 0 | 0 | 1 | 1 | 0 | 1 | 0 | 0 | 5 | 4 | 2 | 0 |
| | Sabouraud | 0 | 3 | 3 | 0 | 1 | 3 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 1 | 3 | 0 |
| <u> </u> | Sabouraud + wheat | 1 | 4 | 4 | 2 | 3 | 4 | 1 | 0 | 0 | 0 | 1 | 1 | 0 | 0 | 1 | 5 | 5 | 3 | 0 |
| Scopulariopsis | Czapek-Dox | 0 | 3 | 3 | 1 | 1 | 3 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 1 | 1 | 1 | 0 |
| brumptii | Czapek-Dox + wheat | 1 | 3 0 | 4 | 2 0 | 5 | 5 | 0 | 0 | 0 | 0 | 1 | 0 | 4 | 0 | 0 | 5 | 4 | 1 | 0 |
| | Sabouraud Sabouraud + wheat | 0 2 | 0 3 | 0 4 | 0 1 | 0 5 | 0 3 | 0 0 | 0 0 | 0 0 | 2 0 | 1 2 | 0 0 | 0 5 | 0 0 | 0 2 | 1 5 | 2 4 | 0 2 | 0 0 |
| | Jabourauu + Wriedl | 2 | S | 4 | 1 | 2 | С | 0 | 0 | 0 | U | Z | 0 | Э | U | Z | Э | 4 | Z | U |

Table 1. Activity of mold exoenzyme isolated from wheat on media with and without addition of wheat

Table 1. Cont.

| Isolate | Medium | | | | | | | | Er | ızym | e act | ivity* | , * * | | | | | | | |
|-------------|--------------------|---|---|---|---|---|---|---|----|------|-------|--------|--------------|----|----|----|----|----|----|----|
| | | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 |
| Stemphylium | Czapek-Dox | 0 | 2 | 4 | 1 | 2 | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 0 | 0 | 1 | 5 | 5 | 0 | 0 |
| sp. | Czapek-Dox + wheat | 1 | 0 | 1 | 0 | 1 | 0 | 1 | 1 | 0 | 3 | 1 | 0 | 1 | 0 | 2 | 5 | 5 | 0 | 0 |
| | Sabouraud | 5 | 2 | 1 | 0 | 2 | 2 | 0 | 1 | 0 | 5 | 3 | 1 | 0 | 1 | 0 | 5 | 4 | 0 | 0 |
| | Sabouraud + wheat | 4 | 2 | 1 | 0 | 3 | 2 | 0 | 2 | 1 | 3 | 2 | 0 | 3 | 1 | 3 | 5 | 5 | 0 | 0 |

*Enzymes: 1) alkaline phosphatase, 2) esterase (C4), 3) esterase lipase (C8), 4) lipase (C14), 5) leucine arylamidase, 6) valine arylamidase, 7) cystine arylamidase, 8) trypsin, 9) α -chymotrypsin, 10) acid phosphatase, 11) naphthol-as-bi-phosphohydrolase, 12) α -galactosidase, 13) β -galactosidase, 14) β -glucuronidase, 15) α -glucosidase, 16) β -glucosidase, 17) N-acetyl- β -glucosaminidase, 18) α -mannosidase, 19) α -fucosidase. **0 – no activity, 1–5 – activity increases with color intensity (1 low activity, 5 high activity).

| Isolate | | Total enzyme a | ctivity summed | l up | Number of active enzymes | | | | | | | |
|-------------------------------|----------------|-----------------------|----------------|----------------------|--------------------------|-----------------------|-----------|----------------------|--|--|--|--|
| | Czapek- Dox | Czapek-Dox + wheat | Sabouraud | Sabouraud + wheat | Czapek- Dox | Czapek-Dox + wheat | Sabouraud | Sabouraud + wheat | | | | |
| Alternaria alternata | 28 | 49** | 23 | 50*** | 12 | 13 | 13 | 14 | | | | |
| Aspergillus fumigatus | 40 | 35 | 41 | 41 | 17 | 10 | 15 | 16 | | | | |
| Aspergillus glaucus | 37* | 23 | 63*^^ | 39^ | 12 | 11 | 16 | 14 | | | | |
| Aspergillus sydowii | 37 | 46 | 35 | 37 | 15 | 14 | 17 | 14 | | | | |
| Aspergillus tamarii | 18 | 21 | 23 | 33^ | 11 | 11 | 12 | 13 | | | | |
| Aspergillus terreus | 14 | 40** | 21 | 38* | 7 | 13 | 11 | 14 | | | | |
| Aspergillus versicolor | 16 | 11 | 27^ | 14 | 6 | 6 | 10 | 7 | | | | |
| Bipolaris holmi | 30 | 49* | 54^ | 56 | 12 | 18 | 16 | 17 | | | | |
| Exserohilum sp. | 54 | 33 | 58* | 41 | 18 | 14 | 18 | 15 | | | | |
| Fusarium proliferatum | 23 | 44**^ | 24 | 32 | 12 | 15 | 10 | 12 | | | | |
| Fusarium tricinctum | 15 | 35** | 29^ | 43 | 8 | 12 | 11 | 11 | | | | |
| Penicillium decumbens | 43^ | 43 | 23 | 57**^^ | 12 | 13 | 7 | 15 | | | | |
| Penicillium expansum | 23^^ | 35* | 8 | 51***^^ | 11 | 12 | 5 | 15 | | | | |
| Scopulariopsis brevicaulis | 22 | 34 | 16 | 35** | 9 | 13 | 8 | 13 | | | | |
| Scopulariopsis brumptii | 16 | 35* | 6 | 38** | 10 | 11 | 4 | 12 | | | | |
| Stemphylium sp. | 25 | 22 | 32 | 37^^ | 12 | 11 | 12 | 14 | | | | |
| Mean | 27.6 | 34.7 | 30.2 | 40.1 | 11.5 | 12.3 | 11.6 | 13.5 | | | | |

| Table 2. Enzyme activity in isola | lates from wheat crop | 5 |
|-----------------------------------|-----------------------|---|
|-----------------------------------|-----------------------|---|

Mann-Whitney-Wilcoxon test for the difference of enzyme activity was performed. Asterisks (*) are stated at the higher number to show the significant difference between pure and supplemented medium and carets (^) are stated at the higher number to show the difference between Czapek-Dox and Sabouraud media (pure and supplemented). The significance of differences: *(^) p < 0.05; **(^^) p < 0.01; ***(^^) p < 0.01.

| Isolate | Medium | | | | | | | | E | nzym | ne act | ivity | ty*,** | | | | | | | | | |
|--------------|------------------|---|---|---|---|---|---|---|---|------|--------|-------|--------|----|----|----|----|----|----|----|--|--|
| | | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 | | |
| Alternaria | Czapek-Dox | 0 | 2 | 4 | 0 | 2 | 2 | 0 | 3 | 0 | 1 | 1 | 1 | 1 | 0 | 2 | 4 | 5 | 0 | 0 | | |
| alternata | Czapek-Dox + rye | 5 | 3 | 4 | 0 | 5 | 3 | 1 | 4 | 0 | 3 | 3 | 4 | 0 | 0 | 4 | 5 | 5 | 0 | 0 | | |
| | Sabouraud | 2 | 2 | 1 | 0 | 2 | 1 | 0 | 0 | 0 | 1 | 2 | 2 | 1 | 0 | 2 | 3 | 3 | 1 | 0 | | |
| | Sabouraud + rye | 4 | 3 | 3 | 2 | 5 | 3 | 0 | 2 | 0 | 3 | 3 | 3 | 5 | 0 | 4 | 5 | 5 | 0 | 0 | | |
| Aspergillus | Czapek-Dox | 4 | 1 | 2 | 0 | 1 | 1 | 0 | 0 | 0 | 4 | 2 | 5 | 3 | 0 | 0 | 3 | 3 | 0 | 0 | | |
| versicolor | Czapek-Dox + rye | 4 | 3 | 4 | 0 | 4 | 5 | 1 | 0 | 1 | 5 | 4 | 5 | 5 | 0 | 1 | 5 | 5 | 1 | 0 | | |
| | Sabouraud | 4 | 2 | 2 | 0 | 1 | 2 | 1 | 0 | 0 | 1 | 2 | 1 | 1 | 0 | 1 | 5 | 2 | 2 | 0 | | |
| | Sabouraud + rye | 5 | 4 | 4 | 1 | 4 | 5 | 1 | 0 | 1 | 4 | 2 | 5 | 5 | 0 | 4 | 5 | 4 | 4 | 0 | | |
| Cladosporium | Czapek-Dox | 2 | 2 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 3 | 0 | 0 | 0 | 4 | 4 | 0 | 0 | | |
| herbarum | Czapek-Dox + rye | 5 | 2 | 4 | 0 | 5 | 4 | 0 | 0 | 0 | 5 | 4 | 5 | 0 | 4 | 2 | 5 | 5 | 4 | 3 | | |
| | Sabouraud | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 3 | 0 | 0 | | |
| | Sabouraud + rye | 5 | 3 | 4 | 0 | 3 | 3 | 1 | 1 | 1 | 5 | 5 | 5 | 0 | 5 | 1 | 5 | 4 | 5 | 5 | | |
| Fusarium | Czapek-Dox | 1 | 2 | 2 | 0 | 1 | 0 | 0 | 4 | 0 | 1 | 1 | 0 | 1 | 0 | 0 | 5 | 0 | 0 | 0 | | |
| cerealis | Czapek-Dox + rye | 5 | 3 | 2 | 0 | 4 | 3 | 0 | 1 | 1 | 5 | 4 | 3 | 2 | 0 | 1 | 5 | 0 | 1 | 0 | | |
| | Sabouraud | 2 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 2 | 0 | 0 | 0 | 0 | 3 | 4 | 0 | 0 | | |
| | Sabouraud + rye | 4 | 2 | 1 | 0 | 3 | 2 | 0 | 0 | 0 | 5 | 3 | 1 | 0 | 0 | 2 | 4 | 4 | 3 | 0 | | |
| Fusarium | Czapek-Dox | 0 | 1 | 2 | 1 | 3 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 2 | 2 | 0 | 0 | | |
| oxysporum | Czapek-Dox + rye | 3 | 3 | 2 | 0 | 4 | 4 | 1 | 4 | 1 | 5 | 5 | 1 | 3 | 0 | 0 | 5 | 0 | 2 | 0 | | |
| | Sabouraud | 3 | 1 | 1 | 1 | 1 | 0 | 0 | 1 | 0 | 3 | 2 | 0 | 0 | 0 | 0 | 5 | 5 | 0 | 0 | | |
| | Sabouraud + rye | 4 | 2 | 3 | 0 | 4 | 3 | 0 | 4 | 0 | 5 | 3 | 0 | 4 | 0 | 1 | 5 | 0 | 1 | 0 | | |
| Fusarium | Czapek-Dox | 2 | 2 | 2 | 0 | 1 | 0 | 0 | 1 | 0 | 4 | 0 | 0 | 0 | 0 | 0 | 4 | 0 | 0 | 0 | | |
| tricinctum | Czapek-Dox + rye | 4 | 1 | 1 | 0 | 3 | 2 | 0 | 1 | 0 | 4 | 4 | 0 | 2 | 0 | 0 | 3 | 0 | 0 | 0 | | |
| | Sabouraud | 3 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 2 | 4 | 1 | 0 | 0 | 0 | 3 | 3 | 0 | 0 | | |
| | Sabouraud + rye | 5 | 3 | 2 | 0 | 3 | 1 | 0 | 0 | 0 | 5 | 5 | 3 | 2 | 0 | 2 | 4 | 5 | 2 | 0 | | |
| Penicillium | Czapek-Dox | 0 | 1 | 1 | 0 | 5 | 2 | 0 | 0 | 0 | 0 | 1 | 0 | 2 | 0 | 3 | 4 | 4 | 0 | 2 | | |
| chrysogenum | Czapek-Dox + rye | 5 | 4 | 2 | 0 | 5 | 4 | 1 | 0 | 5 | 5 | 5 | 5 | 4 | 0 | 5 | 5 | 4 | 3 | 3 | | |
| , . | Sabouraud | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 0 | 2 | 0 | 1 | 3 | 1 | 0 | 0 | | |
| | Sabouraud + rye | 4 | 2 | 3 | 0 | 5 | 3 | 0 | 0 | 3 | 5 | 5 | 5 | 4 | 0 | 5 | 5 | 4 | 4 | 4 | | |
| Penicillium | Czapek-Dox | 4 | 0 | 0 | 0 | 1 | 4 | 0 | 0 | 0 | 3 | 2 | 0 | 0 | 0 | 0 | 5 | 4 | 0 | 0 | | |
| diverse | Czapek-Dox + rye | 5 | 3 | 4 | 0 | 5 | 4 | 1 | 0 | 1 | 5 | 5 | 4 | 3 | 2 | 3 | 3 | 4 | 4 | 0 | | |
| | Sabouraud | 2 | 0 | 0 | 0 | 3 | 0 | 0 | 0 | 0 | 1 | 2 | 0 | 0 | 0 | 0 | 3 | 3 | 0 | 0 | | |
| | Sabouraud + rye | 4 | 2 | 1 | 0 | 5 | 1 | 0 | 0 | 0 | 5 | 5 | 5 | 3 | 0 | 4 | 5 | 4 | 5 | 0 | | |
| Rhizopus | Czapek-Dox | 2 | 4 | 3 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 0 | 0 | 1 | 2 | 1 | 0 | | |
| oryzae | Czapek-Dox + rye | 5 | 2 | 3 | 0 | 3 | 3 | 0 | 0 | 0 | 5 | 5 | 4 | 5 | 0 | 0 | 5 | 1 | 2 | 0 | | |
| | Sabouraud | 4 | 3 | 3 | 0 | 2 | 3 | 2 | 0 | 0 | 3 | 3 | 2 | 0 | 0 | 0 | 4 | 0 | 2 | 0 | | |
| | Sabouraud + rye | 5 | 3 | 3 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 2 | 5 | 3 | 0 | 0 | 5 | 5 | 3 | 0 | | |
| Ulocladium | Czapek-Dox | 1 | 3 | 4 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 3 | 0 | 0 | 0 | 1 | 4 | 4 | 0 | 0 | | |
| chartarum | Czapek-Dox + rye | 2 | 2 | 2 | 0 | 3 | 3 | 1 | 1 | 0 | 1 | 1 | 2 | 4 | 3 | 5 | 5 | 5 | 0 | 0 | | |
| | Sabouraud | 3 | 3 | 1 | 0 | 4 | 3 | 0 | 0 | 0 | 2 | 1 | 3 | 1 | 1 | 3 | 5 | 5 | 0 | 0 | | |
| | Sabouraud + rye | 5 | 4 | 0 | 0 | 5 | 4 | 1 | 0 | 0 | 4 | 3 | 3 | 5 | 3 | 4 | 5 | 5 | 1 | 0 | | |

Table 3. Activity of mold exoenzyme isolated from rye on media with and without addition of rye

*Enzymes: 1) alkaline phosphatase, 2) esterase (C4), 3) esterase lipase (C8), 4) lipase (C14), 5) leucine arylamidase, 6) valine arylamidase, 7) cystine arylamidase, 8) trypsin, 9) α -chymotrypsin, 10) acid phosphatase, 11) naphthol-as-bi-phosphohydrolase, 12) α -galactosidase, 13) β -galactosidase, 14) β -glucuronidase, 15) α -glucosidase, 16) β -glucosidase, 17) N-acetyl- β -glucosaminidase, 18) α -mannosidase, 19) α -fucosidase; **0 – no activity, 1–5 – activity increases with color intensity (1 low activity, 5 high activity).

and Sabouraud Dextrose Broth enriched with cereal (wheat or rye). Isolated strains were also classified in the scale of biosafety levels (BSL).

Material and methods

The study used 23 strains of fungi isolated from samples of grain, grain dust obtained during threshing and soil from the crops of wheat and rye collected in the Lublin area. In order to prepare the isolates for the API ZYM test, the strains were grown initially on two solid media: Malt Agar (Becton, Dickinson and Co.) intended for all kinds of fungi and Potato Dextrose Agar (Becton, Dickinson and Co.) which is a selective medium for *Fusarium* spp. Samples were incubated at 24°C or 30°C for 72 h depending on the medium, and then at room temperature until all have been producing spores. Then the biopsy was taken (5 mm in diameter) from the margin of

| | | Total enzyme ad | tivity summed | up | Number of active enzymes | | | | | | | | |
|----------------------------|----------------|---------------------|---------------|--------------------|--------------------------|----------------------|-----------|--------------------|--|--|--|--|--|
| Isolate | Czapek- Dox | Czapek-Dox + rye | Sabouraud | Sabouraud + rye | Czapek- Dox | Czapek- Dox + rye | Sabouraud | Sabouraud + rye | | | | | |
| Alternaria alternata | 27^ | 48* | 14 | 54** | 11 | 14 | 8 | 16 | | | | | |
| Aspergillus versicolor | 29 | 53** | 27 | 58** | 11 | 15 | 14 | 16 | | | | | |
| Cladosporium herbarum | 18^ | 57** | 6 | 61*** | 8 | 14 | 4 | 17 | | | | | |
| Fusarium cerealis | 18 | 40* | 13 | 34** | 9 | 14 | 6 | 12 | | | | | |
| Fusarium oxysporum | 13 | 43** | 23 | 39* | 8 | 14 | 10 | 12 | | | | | |
| Fusarium tricinctum | 16 | 25 | 18 | 42**^ | 7 | 10 | 8 | 13 | | | | | |
| Penicillium chrysogenum | 25^ | 65** | 9 | 61*** | 10 | 16 | 5 | 15 | | | | | |
| Penicillium diverse | 23 | 56** | 14 | 49** | 7 | 16 | 6 | 13 | | | | | |
| Rhizopus oryzae | 16 | 43* | 31^ | 36 | 9 | 12 | 11 | 11 | | | | | |
| Ulocladium chartarum | 22 | 40 | 35 | 52** | 8 | 15 | 13 | 14 | | | | | |
| Mean | 20.7 | 47.0 | 19.0 | 48.6 | 8.8 | 14.0 | 8.5 | 13.9 | | | | | |

Table 4. Enzyme activity in isolates from rye crops

Mann-Whitney-Wilcoxon test for the difference of enzyme activity was performed. Asterisks (*) are stated at the higher number to show the significant difference between pure and supplemented medium and carets (^) are stated at the higher number to show the difference between Czapek-Dox and Sabouraud media (pure and supplemented). The significance of differences: *(^) p < 0.05; **(^^) p < 0.01; ***(^^) p < 0.01.

the culture and transferred to four liquid media: Czapek-Dox Broth (Becton, Dickinson and Co.) pure and supplemented with 1% of grain (wheat or rye), and Sabouraud Dextrose Broth (Becton, Dickinson and Co.) pure and supplemented with 1% of grain. Media without the tested isolates were used as controls. The isolates were then incubated for 24 days at 24°C. Supernatants obtained from culture (after centrifugation or sedimentation) were tested. Supernatants from control cultures were used as a control. API ZYM tests were performed according to the manufacturer's instructions by placing 65 μ l of the supernatant at appropriate points of the test strip. Reading was made visually. Enzyme activity was determined using a scale of 0 to 5, with 0 indicating a negative reaction. Differences in enzyme activity were compared in cultures with and without addition of the grain extract and statistically tested with Mann-Whitney-Wilcoxon test (R statistical software environment version 3.0.2) [6].

With the help of available literature the tested isolates were classified into classes of biosafety (BSL) [7].

Results

API ZYM test performed on all the control substrates tested showed no activity of hydrolytic enzymes. The following strains isolated from cultures were characterized by the greatest number of active hydrolytic enzymes: Exserohilum sp., Bipolaris holmi (Tables 1 and 2) in strains isolated from samples of wheat and Aspergillus versicolor, Penicillium chrysogenum, Alternaria alternata isolated from samples of rye (Tables 3 and 4). The highest activity of hydrolytic enzymes (4 or 5 in the adopted scale of activity) was found in the strains of Bipolaris holmi and Penicillium decumbens which were grown on media containing 1% of wheat grains (Tables 1 and 2); and in strains of Cladosporium herbarum, Aspergillus versicolor, and Alternaria alternata grown on media containing 1% of rye grains (Tables 3 and 4). In isolates from wheat samples the most active enzymes were N-acetyl- β glucosyloamidase, β-glucosidase, acid phosphatase (Table 1) while in isolates from rye samples, β -glucosidase, N-acetyl- β -glucosyloamidase, and alkaline phosphatase exhibited the highest activity (Table 3).

Comparing the activity of extracellular enzymes in the medium with and without the addition of grain, it was found that supplementation with cereal grain (wheat, rye) resulted in increased activity of the enzymes produced by the isolates in most cases. From 16 isolates of wheat crops in Czapek-Dox medium with an addition of wheat, in 7 cases statistically significant higher enzyme activity was found, and in one case statistically significant lower enzyme activity was found as

compared to pure Czapek-Dox medium. In the Sabouraud medium with an addition of wheat in 6 cases statistically significant higher activity was found, and in 2 cases statistically significant lower activity was detected comparing to pure Sabouraud medium (Tables 1 and 3). The differences in isolates from rye were more distinct than in Czapek-Dox medium with an addition of rye: 8 from 10 isolates showed statistically significant higher activity, and in Sabouraud medium significant higher activity was found in 9 isolates (Tables 3 and 4). When differences between media were compared the differences were much smaller. In wheat isolates higher activity in pure media was found in 4 Sabouraud cultures and in two Czapek-Dox cultures, and in supplemented media 5 Sabouraud cultures showed higher enzyme activity and one Czapek-Dox culture. The differences between media in rye isolates were smaller.

The vast amount of isolates was assigned to BSL-1 (Table 5), i.e. to the class which represents saprophytes or plant pathogens causing a non-invasive or mild threat.

Table 5. Biosafety levels of isolated molds [7]

| No. | Isolate | BSL |
|-----|----------------------------|-----|
| 1 | Alternaria alternata | I |
| 2 | Aspergillus fumigatus | П |
| 3 | Aspergillus glaucus | I |
| 4 | Aspergillus sydowii | I |
| 5 | Aspergillus tamarii | I |
| 6 | Aspergillus terreus | П |
| 7 | Aspergillus versicolor | I |
| 8 | Bipolaris holmi | ND |
| 9 | Cladosporium herbarum | I |
| 10 | Exserohilum sp. | ND |
| 11 | Fusarium cerealis | ND |
| 12 | Fusarium oxysporum | П |
| 13 | Fusarium proliferatum | I |
| 14 | Fusarium tricinctum | ND |
| 15 | Penicillium chrysogenum | I |
| 16 | Penicillium decumbens | I |
| 17 | Penicillium diverse | ND |
| 18 | Penicillium expansum | I |
| 19 | Rhizopus oryzae | I |
| 20 | Scopulariopsis brevicaulis | II |
| 21 | Scopulariopsis brumptii | II |
| 22 | Stemphylium sp. | ND |
| 23 | Ulocladium chartarum | I |

ND – not defined.

Discussion

One of molds, which had high enzymatic activity, belonged to the genus Exserohilum, which consists of more than 30 species. These saprophytes are commonly found on the remains of plants, some of which can cause diseases of plants, animals and humans (immunocompromised persons are particularly at risk) [8]. Another mold showing high activity of the enzymes was species from Bipolaris genus. This type includes more than 100 species, most of them are saprobiotic species found in the soil and some of them have the potential pathogenicity to animals and humans [9]. A significant amount of hydrolytic enzymes was also found in an isolate from the genus Alternaria, the causative agent of phaeohyphomycosis or onychomycosis, sinusitis, ulcerative skin infections, corneal inflammation, and deep mycosis. Infections caused by this genus are a growing problem in patients who have been treated with immunosuppressants. It is also a common allergen [10]. Aspergillus versicolor is described as a rare cause of various mycoses in humans. It is often isolated from fungal diseases of domestic animals and habitats of various soil [3]. The genus Cladosporium is very rarely a cause of opportunistic infections in humans. It is a common allergen [10]. *Cladosporium herbarum* is the most common species in this genus, inhabiting dead organic remains of plants in soil habitats. Molds from the genus Penicillium occur commonly in the fields and meadows, and is also found in flats and basements. It is often present in fruit juices, on citrus and moldy bread. This can cause infections particularly in people with immunodeficiency. High concentration of spores of this fungus in the air can act as an allergen. P. chrysogenum species has a worldwide range. It was isolated many times from soil in different climatic zones, including the polar zone [3].

Hydrolytic enzymes that are released into the environment by dermatophytes, yeasts, and molds are an important element in the pathogenesis of infection [11]. The enzymatic activity of fungi can colonize most surfaces dead or living. The enzymes produced by fungi allow the degradation and the use of most of the naturally occurring compounds and synthetic substances. Decomposition processes are initiated by adaptive enzymes, produced in response to the "signal" from the environment. Changing the proportions of trophic components in the substrate or appearance of trace amounts of a new substrate, stimulates production of specific enzymes [12]. The hydrolases which are synthesized by pathogenic fungi include esterases, and they include the carboxyl group ester hydrolases (triacylglycerol lipase, phospholipase A2), the phosphoric monoester hydrolases (alkaline phosphatase, acid phosphatase) and sulfuric ester hydrolases (sulfatases); glycosidases (α -glucosidase, β -glucosidase, α -mannosidase); peptidases – exopeptidases (proteinases, e.g. aspartic proteinase, aminopeptidases, e.g. arylamidase) and endopeptidase (proteinase hydrolyzing C-N bond, e.g. urease). Using a commercial standardized test (API ZYM, bioMérieux), one can create a different fungal species enzymogram illustrating the characteristic enzymatic activity [3, 13–15].

Obtained results tend to reflect methods of storage and processing of grain; whether the technology completely eliminates molds from food. As the results show, some molds (e.g. *Exserohilum* sp., *Bipolaris holmi, Alternaria alternata, Cladosporium herbarum, Aspergillus versicolor, Penicillium chrysogenum*) isolated from samples of grain, grain dust, and soil from crops of wheat and rye, probably will also be able to show significant enzyme activity in natural environment, which may help to adapt to new conditions, and to colonize various locations and affect human health (especially in immunocompromised persons) exposed to inhalation of large amounts of dust generated during harvesting grain cereals (i.e. threshing with a combine harvester or pouring grain into barns).

Conclusions

Hydrolytic exoenzyme activity in tested fungi depends on the type of the medium and the addition of the grain extract. Particular grains can stimulate activity of extracellular enzymes in tested fungi. Activation of enzymes was influenced by an addition of wheat and rye to the medium. The relationship between high enzymatic activity of examined fungi and BSL scale was not found.

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Conflict of interest

The authors declare no conflict of interest.

References

- 1. Nowicki R, Korting HC. Differences in the hydrolytic activity of dermatophytes [Polish]. Mikol Lek 1995; 2: 209-13.
- 2. Łukaszuk C, Krajewska-Kułak E, Niczyporuk W, et al. Variations of enzymatic activity and biotypes of the yeast like fungi strains isolated from cancer patients. Ann Acad Med Bialost 2005; 50: 16-9.
- 3. Baran E. Outline of medical mycology [Polish]. Volumed, Wroclaw 1998; 24-5, 96, 118, 182, 307, 518-9.
- 4. Information of the Mycological Section Board of Polish Dermatological Society, Department of Dermatology and Venereology, Wroclaw Medical University [Polish]. Mikol Lek 1998; 193-6.
- Pałczyński C, Kieć-Świerczyńska M. Allergology and clinical toxicology. Clinical Occupational Medicine. J. Nofer Institute of Occupational Medicine, Lodz 2000; 47: 73-4.
- 6. R Core Team R. A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria 2013. URL: http://www.R-project.org/.

- 7. de Hoog GS. Risk assessment of fungi reported from humans and animals. Mycoses 1996; 39: 407-17.
- 8. da Cunha KC, Sutton DA, Gené J, et al. Molecular identification and in vitro response to antifungal drugs of clinical isolates of Exserohilum. Antimicrob Agents Chemother 2012; 56: 4951-4.
- 9. da Cunha KC, Sutton DA, Fothergill AW, et al. Diversity of bipolaris species in clinical samples in the United States and their antifungal susceptibility pofiles. J Clin Microbiol 2012; 50: 4061-6.
- 10. Krzyściak P, Skóra M, Macura AB. Atlas of human pathogenic fungi [Polish]. MedPharm Poland 2011; 316.
- 11. Nabrdalik M. Harmfulness of moulds occurring in human environment [Polish]. Mikol Lek 2007; 14: 195-200.
- Dynowska M. Współczesne poglądy na taksonomię, pochodzenie i naturę grzybów. In: Mikologia – co nowego? Baran E (ed.) [Polish]. Cornetis, Wroclaw 2008; 127.
- 13. Krzyściak P. Evaluation of the enzymatic activity of fungi of the genus Rhodotorula isolated from human performed with API ZYM test [Polish]. Mikol Lek 2011; 18: 65-70.
- 14. Kozak M, Sobczak P, Żukiewicz-Sobczak W. Health properties of selected herbal plants. Health Probl Civil 2016; 10: 64-70.
- 15. Walczak J, Pomastowski P, Buszewski B. Food control quality by hyphenated separation techniques. Health Probl Civil 2015; 9: 33-8.